



Euthanasia using gaseous agents in laboratory rodents: how to refine and improve animal welfare and user safety



A bit of etymology



The word 'euthanasia' is derived from the Greek words 'eu' meaning good and 'thanatos' meaning death

A 'good death' would be one that occurs with minimal pain and distress

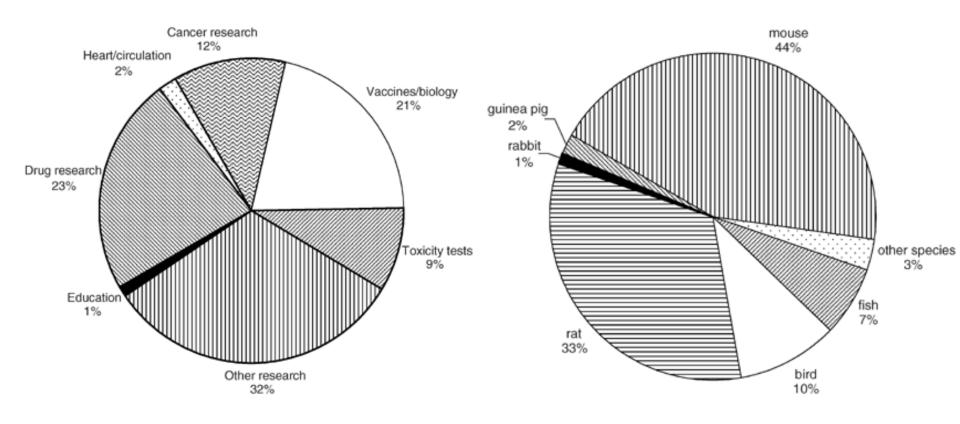
• In the case of animals, the word 'euthanasia' is often substituted by terms such as 'humane death' or 'humane killing'

For simplicity, the term 'euthanasia' will be used throughout this presentation

Why consider euthanasia for use in laboratory rodents?



 Rodents remain the most widely used species for scientific research due to their small size, low cost, rapid sexual maturity and scope for genetic manipulation



Why consider euthanasia for use in laboratory rodents?



- Despite significant interest in the replacement of animals for scientific purposes, the numbers involved are still growing
- Laboratory rodents are euthanized for various reasons

To provide tissues for scientific purposes, at the end of an experiment

When adverse effects (pain, distress, suffering, etc.) become excessive

When animals become unwanted stock

 Regulation requires animals to be euthanized upon completion of the work and when humane endpoints are reached

Identification and use of methods that offer a death with minimal suffering is a moral imperative, and crucial to support the justifiability of animal-based research based on utilitarian harm/benefit ethical reasoning

Overview of euthanasia methods



Overview of euthanasia methods from published euthanasia guidelines for adult laboratory rodents

	UK ⁹	EU ^{12,149}	US ⁶	Canada ⁸	New Zealand ⁷	Australia ⁷
Cervical dislocation Decapitation Concussion by blunt force trauma Exposure to CO ₂ Exposure to CO Overdose of anaesthetic Microwave irradiation						

CO, carbon monoxide, CO₂, carbon dioxide; EU, European Union; UK, United Kingdom; US, United States.

- Methods permitted or recommended for use
- Methods where additional permissions are required
- Methods not permitted or recommended



- ➤ Concern for animal welfare during killing stems from animals' ability to experience potentially negative sensations occurring during the conscious phase of the process
- > To determine the welfare consequences of a euthanasia method

Time taken for the animal to be rendered unconsciousness inclusive of pre-handling time and euthanasia method application

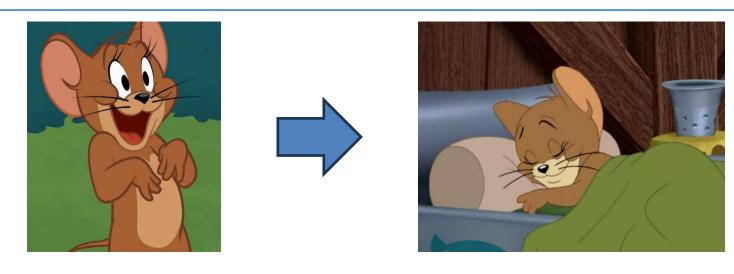
Presence and character of potentially negative experiences during the conscious phase











By assessing the gradual loss of reflexes and goal-directed behaviours and ultimately, motor coordination

Typically measured in rodents by their inability to effectively right their posture, referred to as loss of righting reflex or loss of posture

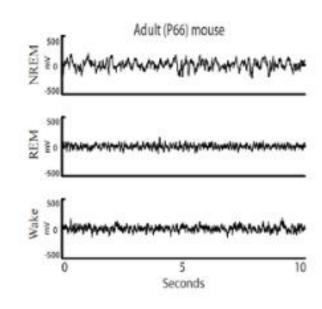




Changes in behavioural output can be measured through the transition from goal directed behaviour (motivated escape behaviours) to the presence of spontaneous ones such as jumping, gasping

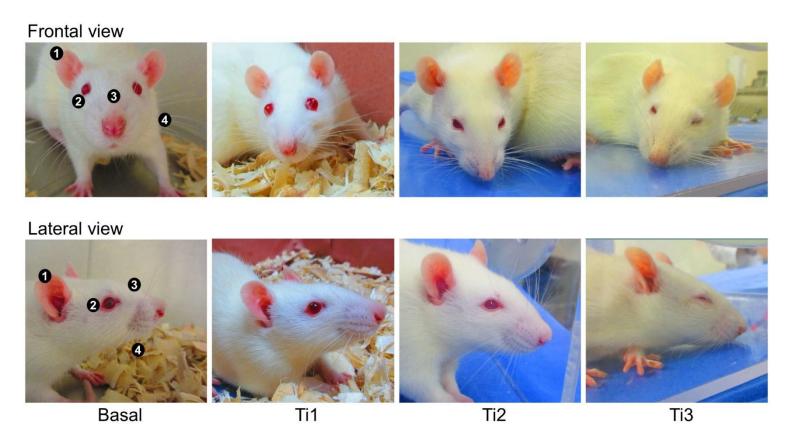


Unconsciousness is also characterised by the presence of high amplitude, low frequency activity in the EEG signal





Presence and character of potentially negative experiences during the conscious phase

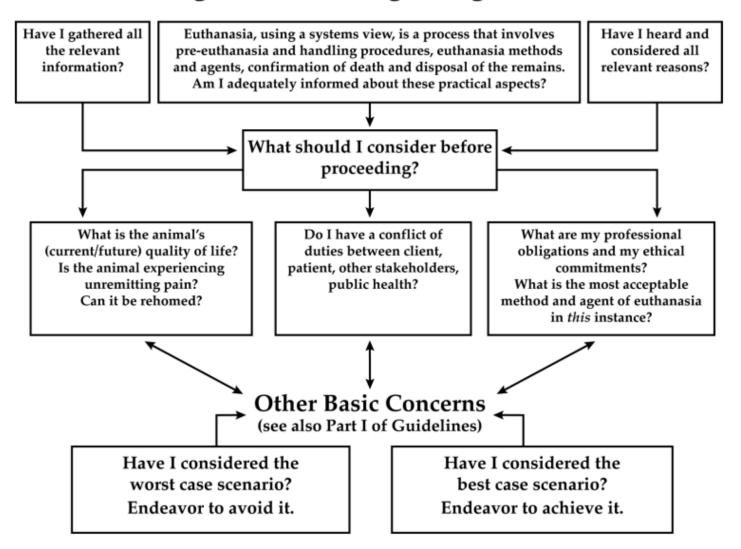


Example of the FAU and the assigned score. (1) ear change; (2) orbital tightening; (3) nose/cheek flattening; and (4) whisker change

Assessing user welfare during euthanasia



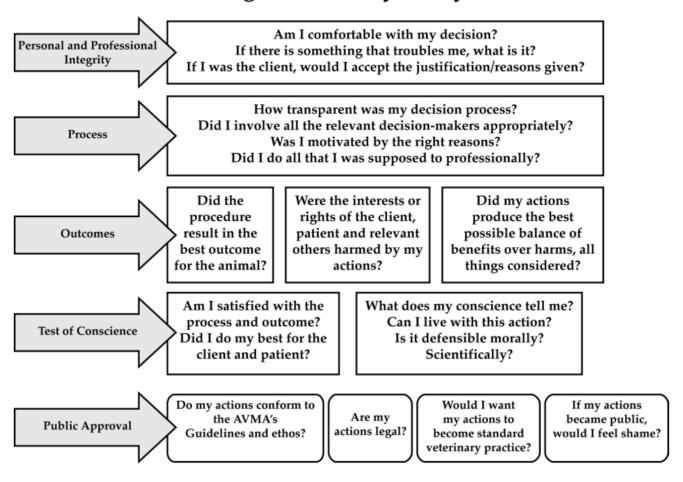
Making a Decision Regarding Euthanasia



Assessing user welfare during euthanasia



Evaluating the Morality of My Decision



Euthanasia using various methods



Agent or Method	2013 AVMA Guidelines ⁶³	2016 Society of Mammologists Guidelines ⁸⁹	2010 CCAC Euthanasia Guidelines ²²	2010 EU Directive ²
Barbiturate	Aa	A ^b	A^a	A ^c
Dissociative Agent Combination	A	A^b	_	A^c
Ethanol	C^d	_	_	_
T-6131	_	_	$C^{e,f}$	A ^c
Carbon Dioxide	$C^{g,k}$	_	$C^{g,h}$	$C^{g,i}$
Carbon Monoxide	C^{l}	_	_	_
Cervical Dislocation	C ^{n,o}	C^p	$C^{n,o,q,r}$	C^s
Decapitation	C^{t}	_	$C^{t,u}$	C^{v}
Inhalant Anesthetic	Cw,x	A^b	$A^{w,y}$	A^c
Focused Beam Microwave Irradiation	C^{a1}	_	_	_
Nitrogen, Argon	U	_	C^{b1}	A
Nitrous oxide	U	_		
Exsanguination	U^{m}	_	U^{m}	U^{m}
Thoracic Compression	U	C ^{c1}	_	_
Blunt Force Trauma to the Head	Cf,d1	_	_	C^{d1}

A = Acceptable; C = Acceptable with Conditions; U = Unacceptable when used as sole agent on conscious animals; - = not addressed.

Euthanasia using various methods



- a, IV preferred over IP; concentrated solutions may cause pain when given IP.
- b, Drug use in field can present additional risks to investigators and stress to animals, risk of secondary toxicity if carcass left in field to be eaten.
- c, Anesthetic overdose should, where appropriate, be used with prior sedation
- d, 0.5 mL of 70% IP for mice; unacceptable for larger species
- e, Only IV, slowly
- f, Personnel must be well-trained
- g, Gradual fill only, displacing 10% to 30% chamber volume per minute; source of gas should be compressed gas cylinder; euthanize in home cage or euthanasia chamber should be emptied and cleaned between uses; verify death has occurred
- h, Must have written SOP, written records, regular postapproval monitoring, animals should be anesthetized prior to CO, delivery
- i, Not to be used on fetuses/neonates
- j, Prefilled chamber recommended for guinea pigs to minimize the experience of breathlessness
- k, Prolonged exposure required for neonates
- l, Requires properly maintained equipment; hazardous to personnel, acceptable only when conditions for safe use can be met
- m, Acceptable under deep anesthesia
- n, Personnel must be trained and their proficiency validated; availability of secondary method if initial attempt unsuccessful
- o, For rodents < 200 g
- p, Animals of small body size, performed by experienced personnel
- q, Anesthetize or sedate first; scientific justification required for use on conscious animals
- r, For rats > 200g use commercial dislocator
- s, Mice, rats < 150 g
- t, Properly maintained equipment: blades sharp, clean, in good condition; operator skilled in handling/restraint of animals; personnel should be trained on dead/anesthetized animals to demonstrate proficiency
- u, Recommend anesthetizing first
- v, Use only if other methods are not possible
- w, Time to death may be prolonged, consider adjunctive method once animals are deeply anesthetized
- x, Maintain compatible groups, clean and maintain induction/euthanasia chamber, adhere to recommended flow rates, it is important to verify death when inhalant method used
- y, Not for use in species that breath-hold
- z, Occupational health and safety issue for personnel exposed to waste anesthetic gas
- a1, Purpose-built equipment, mouse and rat only
- b1, When scientifically justified and approved by ACUC; Argon is aversive to rats; O2 concentration must be <2%, only appropriate for use
- if O2 concentration is known/measured; mixtures of argon and nitrogen should only be used if animal is already anesthetized
- c1, Personnel skilled in technique and animal small enough to allow thoracic cavity to be collapsed and prevent inspiration
- d1, Small laboratory rodents <1kg
- e1, T-61 not available in the US

CO₂ as an euthanasia method



- Carbon dioxide narcosis and asphyxiation have long been used for euthanasia of rodents and other laboratory species
- At present, the method is considered "acceptable with conditions" by the AVMA

Specified displacement rate

Prefilled chambers are unacceptable

Verify that an animal is dead after exposure to CO₂ as with other inhalant euthanasia techniques

In the past, CO₂ was administered by putting animals in a prefilled chamber or delivering
 CO₂ at a very high rate (approximately 70%) of volume displacement

CO₂ as an euthanasia method



- Cost-effective
- Relatively safe for users and the environment Suitable for euthanasia of multiple rodents at the same time
- Anesthetic at high concentrations (30% to 40% in rats)
 - Renders animals unconscious before they die of respiratory arrest and hypoxia

- Reaction with the fluid in mucous membranes to form carbonic acid
- Produces a stinging sensation in the eyes and throat
 - Produces anxiety responses in rodents at concentrations above 20%

Controversies regarding its use, leading to find alternatives to CO₂ use



- The use of CO₂ for euthanizing animals has been questioned as studies have shown that the exposure to CO₂ causes aversion in rats and mice, demonstrated by several behavioral tests
- Assessment and development of alternatives to CO₂ has become a focal point
- Recent studies recommend the use of volatile anesthetics, but it still needs to be clarified whether they cause less pain, distress, and suffering compared to CO₂

Euthanasia of laboratory mice: Are isoflurane and sevoflurane real alternatives to carbon dioxide?

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- The appropriate surrogate measure for unconsciousness in rodents is not finally defined
- The loss of righting reflex (LORR) has been described as a first indicator of insensibility and as a good correlate for the loss of consciousness in humans



C57Bl/6J	LORR [s]		
CO ₂ 20	108.0	[84.5/117.5]	
CO ₂ 60	60.0	[57.0/63.75]	
CO ₂ 100	50.0	[49.0/52.5]	
Iso 2%	122.0	[108.0/136.0]*+	
Iso 5%	67.0	[65.5/68.0] ^{+#}	
Sevo 4.8%	122.0	[113.25/129.5]	
Sevo 8%	82.0	[75.5/86.0]*+	



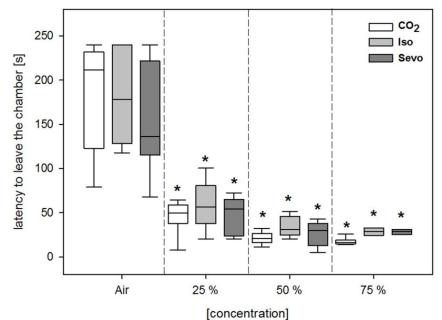
Set-up of the approachavoidance test

Which gas is more aversive to animals?



- CO₂, isoflurane and sevoflurane were perceived similar aversive by the animals
- Indicated by shorter escape latencies compared to Air control

Difficult to weigh up the advantages and disadvantages of CO₂





Data does not conclusively demonstrate the advantages of halogenated over CO₂



> So why not try to improve the existing methods? Which will lead to improved animal welfare



Know the basics : Gas source

- Carbon dioxide and CO₂ gas mixtures must be supplied in a precisely regulated and purified form without contaminants
 - > Commercially supplied cylinder or tank
- Gas displacement rate is critical to the humane application of CO₂
 - Appropriate pressure-reducing regulator and flow meter with demonstrated capability for generating the recommended displacement rates for the size container being utilized







Know the basics : Animal age is crucial

On the day of birth, rats and mice exposed to 100% CO₂ required exposure times of 35 and 50 minutes, respectively, to ensure death

➤ If exposure time is not sufficient, pup will recover as shown by its pink coloration



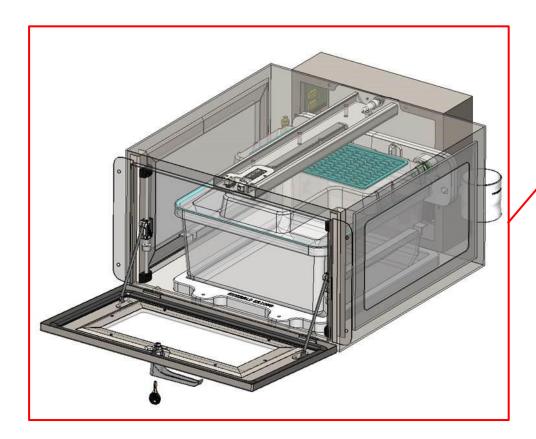
Rat and mouse pups are born neurologically immature when compared with humans, and their afferent pain pathways are not well developed until after postnatal day 5 to 7







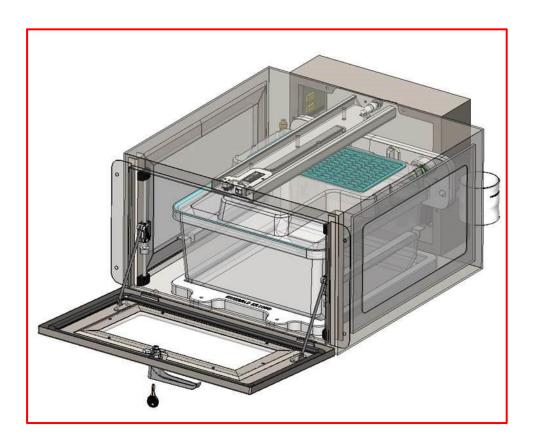
 AVMA Guidelines on Euthanasia state that, to decrease potential distress of animals, the home cage should be used for the euthanasia of mice







 AVMA Guidelines on Euthanasia state that, to decrease potential distress of animals, the home cage should be used for the euthanasia of mice

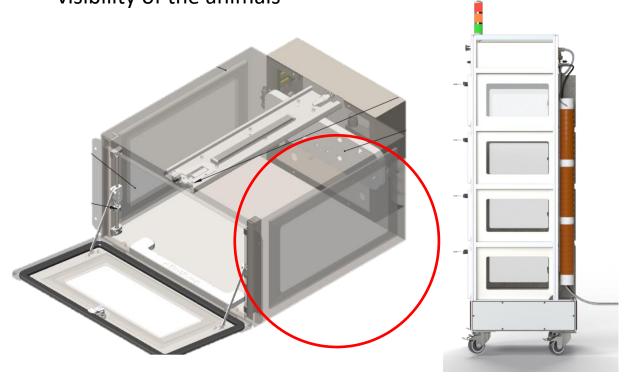


Offers several advantages over physical methods

- Non-contact nature
- Elimination of stress associated with handling
- Isolation and restraint
- Minimising the impact of operator error



The euthanasia system should allow easy visibility of the animals



- Windows from each side of the system allowing optimal visibility
 - > Red tainted windows as an option





Appropriate gradual displacement rate

Displacement rate	Time to unconsciousness	CO ₂ concentration	
20% / min	106 s	30 %	
10% / min	156 s	21 %	
50% / min	< 80 s	45 %	

➤ CO₂ exposure using a gradual-fill method is less likely to cause pain prior to unconsciousness due to nociceptor activation by carbonic acid



AVMA Guidelines on Euthanasia recommends an optimal flow rate for CO₂ euthanasia systems that can displace 30% to 70% of the chamber or cage volume/min

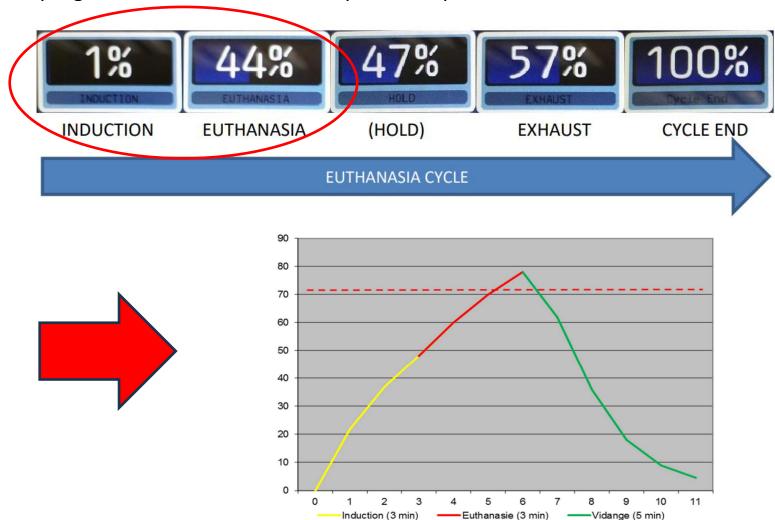


- Our system is equipped with independent modules
- > Each module can be programmed with specifics flow rate
- Can evolve with AVMA and others guidelines



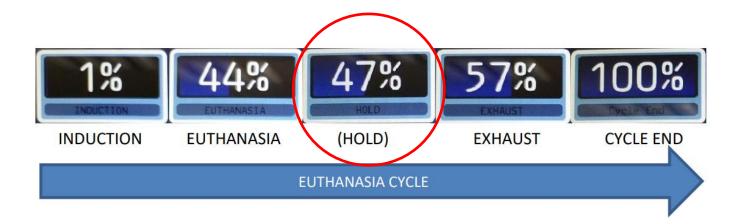


Each program should include these specific steps :





 Whenever gradual displacement methods are used, CO₂ flow should be maintained for at least 1 minute after respiratory arrest



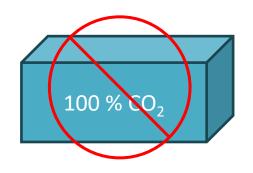
Our system allows the admin to increase or decrease the duration of each phase



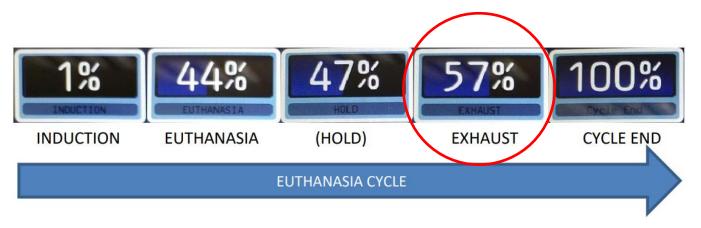


What about prefilled chambers?

Prefilled chambers are not recommended due to the potential for significant pain upon inhalation of the gas



Our system is equipped with an exhaust phase allowing CO₂ to be flushed out of the cage (if the latter is to be reused)



➤ If euthanasia is not conducted in the home cage, induction chambers should be emptied and cleaned between uses



More advices

- The practice of immersion, where conscious rodents are placed directly into a container prefilled with 100% CO₂, is unacceptable
 - A 2-step process, where animals are first rendered unconscious and then immersed, is preferred when gradual displacement methods cannot be used
- Consideration should be given to the benefits of using a darkened home cage, while also keeping in mind the need to have the animal under observation
- If animals need to be combined, they should be of the same species and, if needed, restrained so that they will not hurt themselves or others
- Addition of O₂ to CO₂ will prolong the time to death and may complicate determination of consciousness
 - There appears to be no advantage to combining O₂ with CO₂ for euthanasia

How to increase animal welfare during ${\rm CO_2}$ exposure

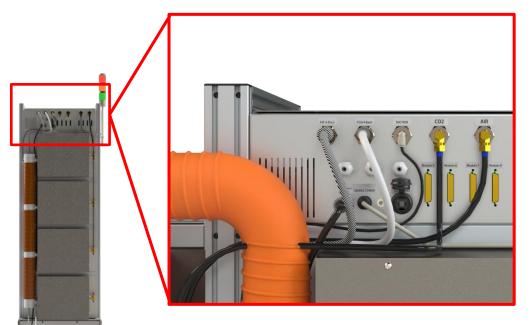


More advices

Your euthanasia equipment has to be equipped with a CO₂ flow alarm



CO₂ room level alarm







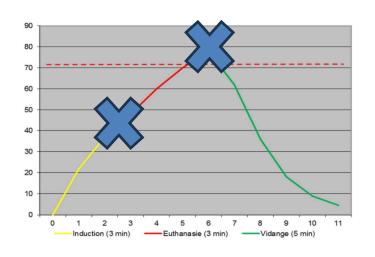


More advices

Your euthanasia equipment has to be equipped with a CO₂ flow alarm



CO₂ room level alarm







What about user safety?

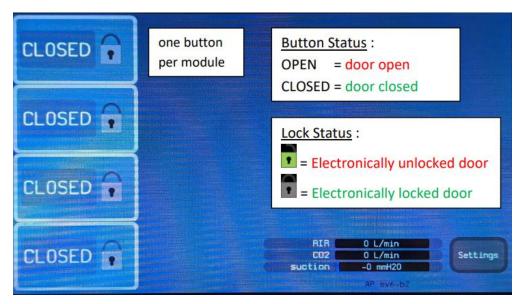
1



Password-equipped system for admin (can change cycle parameters) and users (can only use the system as set by the admin)

Each module will be locked automatically when closed to allow for the cycle launch

Modules can't be opened by user during cycle (a safety feature allows for emergency opening)





What about user safety?

3



System equipped with a signal tower showing the cycle status:

Green: system ready to be used

Orange: system in use, CO₂ is flowing to the cages

Red: system is broken and needs assistance

Signal tower can be seen from distance



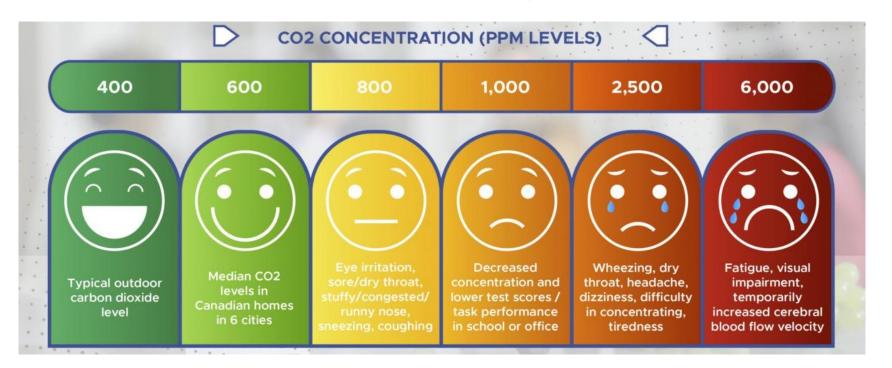
System equipped with an exhaust cycle to filter CO₂



4



System equipped with an exhaust cycle to filter CO₂





A CO₂ room alarm is required in the experimental area

Has to be set with a minimum value of 5000 ppm (0,5%) with sound alarm

Has to be set with a maximum value of 30.000 ppm (3%) with sound and light alarm



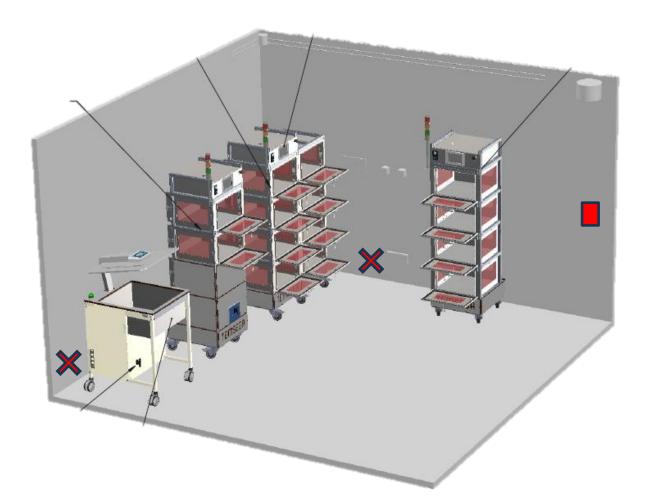


 \triangleright If the sensor has to be installed in a place without ventilation, where CO_2 bottles are kept:

Alarm installed at a height of 30cm above the floor and very close to the potential leak source



- CO₂ is 1.5x heavier than air
- Alarm has to be installed away from air vents, 1.5m above the floor and 1m from any system



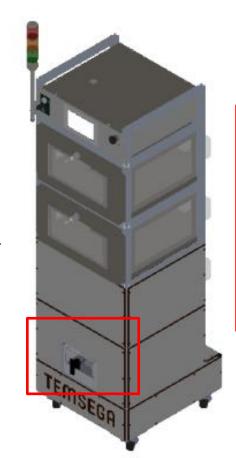


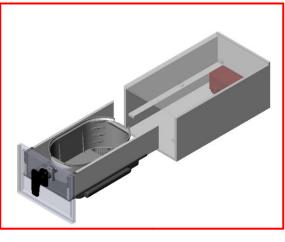
4



System equipped with an exhaust cycle to filter CO₂

Can be equipped with soda lime filter











4



System equipped with an exhaust cycle to filter CO₂

Can be connected to building exhaust (active or passive)







How to increase animal welfare during ${\bf CO_2}$ exposure



> Evolution capability



2 modules 4 modules 6 modules



- Euthanasia system compatible with several cage model from different manufacturers
- A specific adaptation can be made on request

